Assessing Viral Shedding and Infectivity of Tears in Coronavirus Disease 2019 (COVID-19) Patients

Ivan Seah Yu Jun, MBBS, Danielle E. Anderson, PhD, Adrian Eng Zheng Kang, BSc, Lin-Fa Wang, PhD, Pooja Rao, MBBS, Barnaby Edward Young, MB, BChir, David Chien Lye, MBBS, Rupesh Agrawal, MD

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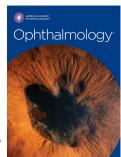
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2	Title: Assessing Viral Shedding and Infectivity of Tears in Coronavirus Disease 2019
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4	Authors (In Order): Ivan Seah Yu Jun, MBBS ¹ ; Danielle E. Anderson, PhD ² ; Adrian Eng Zheng
5	Kang, BSc ² ; Lin-Fa Wang, PhD ² ; Pooja Rao, MBBS ⁴ , Barnaby Edward Young, MB, BChir ^{3, 4, 5} ;
6	David Chien Lye, MBBS ^{3, 4, 5, 6} ; Rupesh Agrawal, MD ^{2, 4, 5, 6, 7}
7	Affiliations
8	¹ National University Hospital, Singapore
9	² Duke-NUS Medical School, Singapore
10	³ National Centre for Infectious Diseases, Singapore
11	⁴ Tan Tock Seng Hospital, Singapore
12	⁵ Lee Kong Chian School of Medicine, Singapore
13	⁶ Yong Loo Lin School of Medicine, Singapore
14	⁷ Moorfields Eye Hospital, NHS Foundation Trust, London, United Kingdom
15	Corresponding Author
16	Associate Professor (Dr) Rupesh Agrawal
17	Address: 11 Jalan Tan Tock Seng, Singapore 308 433
18	Email: Rupesh_agrawal@ttsh.com.sg
19	Contact Number: +65 9061 3202
20	Fax: + 65 6357 1000
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low risk of ocular transmission.

34	Ocular transmission of COVID-19 is uncertain. 64 tear samples were collected from 17
35	COVID-19 patients between Day 3 to Day 20 from initial symptoms. Neither viral culture nor
36	reverse transcription polymerase chain reaction (RT-PCR) detected the virus, suggesting a

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The severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) has rapidly spread across the globe to cause a pandemic. While it is known to be transmitted via droplets, alternative modes of transmission remain unknown. Transmission through infected ocular tissue or fluid has been a controversy^{1,2}. It is hypothesized that the nasolacrimal system can act as a conduit for viruses to travel from the upper respiratory tract to the eye. Hence, ocular tissue and fluid may represent a potential source of SARS-CoV-2. In this study, we attempted to determine the possibility of transmission through tears by assessing for the presence of SARS-CoV-2 with viral isolation and quantitative reverse transcription polymerase chain reaction (RT-PCR). As patients were being monitored clinically via routine nasopharyngeal swabs (NP), they were compared with tears to further understand patterns of viral shedding. 17 COVID-19 patients were recruited for this prospective study in Singapore after obtaining informed consent. This study was carried out in accord with the Declaration of Helsinki and ethics approved by the Domain Specific Review Board of the National Healthcare Group (NHG) Singapore. NPs were collected routinely for clinical monitoring of patient's condition while tear samples were collected purely for research purposes. On some days, both tears and NPs were collected at the same time. These samples were delivered to different labs for processing. COVID-19 patients were tested positive by RT-PCR of NPs in a clinical diagnostic laboratory. NPs were collected in universal viral transport media and RNA extraction done using NucliSENS® easyMAG® system (bioMérieux). 55μl of the elute was then used to perform RT-PCR as per manufacturer's instructions using the A*STAR FORTITUDE kit (Accelerate

Technologies Pte .Ltd, Singapore). The limit of detection was estimated to be <25copies of 62 63 RNA. 64 Tears were sampled by a senior consultant ophthalmologist using Schirmer's test strip at varying timepoints between Day 3 and 20 after the initial development of symptoms. 65 Caution was taken to prevent contamination of samples. The Schirmer's strip tear collection 66 method was previously validated in other studies³. Samples from both eyes were taken and 67 analysed separately. Collected strips were placed into individual falcon tubes of universal 68 69 viral transport media. Samples were delivered to a research laboratory for processing. Samples were used to inoculate Vero-E6 cells (ATCC®CRL-1586TM). After 4 days of 70 incubation, cells were observed for the presence of cytopathic effect (CPE). Total RNA was 71 extracted from all samples using E.Z.N.A. Total RNA Kit I (Omega Bio-tek) according to the 72 manufacturer's instructions and samples were analysed by real-time quantitative reverse 73 transcription-PCR (RT-qPCR) for the detection of SARS-CoV-2 as previously described⁴. 74 75 Clinical data including age, sex, symptoms, nasopharyngeal swab results were collected 76 from electronic health records and correlated with RT-PCR results. Ocular symptoms which were assessed include red eye, tearing, blurring of vision, discharge and colour desaturation. 77 These symptoms were chosen based on the ocular manifestations of other coronaviruses 78 known to infect humans and animals². Other symptoms of COVID-19 assessed include fever, 79 80 cough, shortness of breath, rhinorrhea and sore throat. Of the 17 patients recruited, none presented with ocular symptoms. However, 1 patient 81 82 developed conjunctival injection and chemosis during the stay in the hospital (Table 1 available at www.aaojournal.org). 14 patients presented with upper respiratory tract 83 84 symptoms including cough, rhinorrhea and sore throat.

A total of 64 samples were taken over the study period, with 12, 28 and 24 samples taken 85 from first, second and third week of initial symptoms respectively. All were tested negative 86 for the SARS-CoV-2 on viral isolation and RT-PCR. Tear results were compared with NP 87 results as shown in **Figure 1**. Ct values of NP swabs were featured. 88 To our knowledge, this is the first study comparing viral shedding in tears with NP results 89 during the course of COVID-19 infection. A previous study showed positive SARS-CoV-2 RT-90 PCR results from a patient's tears, but isolation of the virus was unsuccessful⁵. In this study, 91 92 there was no evidence of SARS-CoV-2 shedding in tears through the course of the disease. Viral load detected in nasal and throat swabs are elevated for a period of approximately 2 93 weeks from the onset of COVID-19 symptoms⁶. In this study, the tear sampling timepoints 94 cover these 2 weeks of active infection, providing a good representation of the full disease 95 course. All tear samples tested negative even when NPs continued to test positive. 96 Furthermore, patients with symptoms of upper respiratory tract infections did not 97 98 demonstrate any viral shedding in tears, suggesting the hypothesis of the lacrimal duct as a viral conduit may not be true. Most importantly, only one patient developed ocular 99 symptoms during the disease course and no evidence of SARS-CoV-2 could be found in the 100 tear samples. This suggests that transmission through tears regardless of the phase of 101 infection is likely to be low. 102 The study had several limitations. Firstly, the samples were analysed in different 103 104 laboratories utilising two different assays. As the NPs were utilised in the clinical setting to monitor disease progression, they were analysed in a clinical diagnostics lab while the tear 105 samples were analysed in a research lab. While the limit of detection for the research lab 106 was not assessed due to logistical limitations, it should be noted that the tear samples were 107

incubated with Vero-E6 cells for 4 days prior to obtaining the RNA for RT-PCR. If SARS-CoV-2 existed in the samples, CPE would have been observed even in a false negative RT-PCR result. We observed neither CPE nor a positive RT-PCR result, thereby the likelihood of SARS-CoV-2 being found in the tear samples is still low. Secondly, only tears were sampled rather than conjunctival tissue. In the pandemic setting, COVID-19 patients are already emotionally distraught with their diagnosis. Hence, conjunctival tissue sampling was avoided to reduce patient distress. Despite this, we believe that our results do highlight a low risk of ocular transmission. In the acute infection of conjunctival cells, cells die through viralmediated lysis or from immune reactions. Cell death will release viral material into tears which can still be detected via RT-PCR. Thirdly, the study had a small sample size due to the logistical limitations of the outbreak response. These patients also usually present a few days after symptom development, making sampling during early infection difficult. Finally, only 1 patient had ocular symptoms in our study. However, studying patients with ocular symptoms can be difficult. In a study of 1099 COVID-19 patients, only 0.8% developed conjunctival congestion⁷. The results from this study suggests that the risk of SARS-CoV-2 transmission through tears is low. However, further definitive mechanistic studies are required. SARS-CoV-2 has been known to infect cells via ACE2 receptors. More studies are required to definitely prove the presence of ACE2 on corneal and conjunctival cells. Future studies involving more patients with ocular symptoms should also be considered. Finally, future studies should consider the association between serum viral load and viral shedding in tears. Unfortunately, no blood samples were analysed for this experiment as they were not routine clinical investigation in

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149 Figure Legends

- Figure 1: Comparison of Tear Samples and Nasopharyngeal Sawb Samples Over Course of COVID-19 Illness
- 152 CT results of all nasopharyngeal swabs are displayed. All tear samples were tested 153 neagtive for on both viral isolation & RT-PCR. These results were labelled by a red 154 coloured box.

										Days Sind	e Initial Co	OVID-19 Sy	mptoms									
		Day 1 Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10	Day 11	Day 12	Day 13	Day 14	Day 15	Day 16	Day 17	Day 18	Day 19	Day 20	Discharge Status	Total Duration of Symptoms (As of 12/3/2
	1	24.4	20.11	19.09	NA	21.65	24.00	23.46	23.17												Still admitted	6 Days
	2	24.3	37.92	NA	NA	32.75	-	-													D9 Discharge	5 Days
	3		27.06	22.28	NA.	22.41	24.30	28.50	30.31	26.67	25.80	27.99	38.05	-							Still admitted	5 Days
	4				21.20	22.04	NA	NA	24.51	28.4	28.24	NA	30.54	NA	34.86	NA	36.83	NA	37.17	NA	Still admitted	18 Days
	5				29.48	26.19	NA	day 8	28.69	NA	NA	NA	NA	34.07	NA	NA		35.48 *	NA *	NA *	Still admitted	Still Symptomatic (23 Day
Number	6	33.5 -	31.18	-	36.28	-	-														D8 Discharge	2 Days
	7					37.70	NA	35.02	NA	34.69	35.09	NA	-	-							D15 Discharge	11 Days
Z	8							26.33	NA	NA	NA	NA	29.15	NA	37.05	NA	37.1	NA	35.35	NA	Still admitted	18 Days
500	9							31.22	33.71	NA	34.17	NA	NA	34.63	NA	NA	-	34.25	29.04	35.33	Still admitted	11 Days
5	10					-	-	-	NA	34.10	-										D14 Discharge	12 Days
<u>.</u>	11							29.19	NA	NA	33.72	36.20	NA	36.71	NA	NA	33.13	NA	35.14	NA	Still admitted	15 Days
Patient	12							37.55	32.79	33.43	38.16	32.28	39.39	35.91	37.72	38.42	38.21	-	38.21	36	D25 Discharge	23 Days
	13										29.54	NA	NA	33.04	NA	NA	NA	-	37.55	NA	Still admitted	22 Days
	14									29.30	34.00	33.31	28.66	35.27	34.53	30.77	37.45	35.92	32.29	-	Still admitted	6 Days
	15					25.40	NA	NA	NA	26.38	33.06	32.26	30.20	36.21	29.48	-	-				D17 Discharge	11 Days
	16			22.89	NA	22.15	NA	NA	NA	NA	NA	NA	NA	NA	NA	-	38.81	-	36.34	-	D21 Discharge	20 Days
	17				19.35	NA	20.01	NA	21.97	29.06	32.19	32.27	21.31	19.22	34.10	32.05	30.43	-	36.52		D22 Discharge	15 Days
																				Legend		
																				-	Negative Nasopharyngeal Swal	
																				NA	No Nasophayrngeal Swab Taker	-
																				*	Ocular Symptoms	
																					Tears Sampled Negative	
					Figur	e 1: Comp	arison of	Tears Sam	ples and I	Nasophan	ngeal Swa	b Samples	Ct Values C	over Course	of COVID-	9 Illness					Tears Sampled Positive	

